

Research Paper

**IMPACT FACTORS ON DISTRIBUTION AND CHARACTERISTICS OF
NATURAL PLANT COMMUNITIES IN JAZAN REGION, SAUDI ARABIA**

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Abstract

To identify impact factors on distribution and characteristics of natural plant communities in Jazan region, Saudi Arabia. The main aim of this study was to examine the spatial pattern of vegetation and relationship between vegetation composition and environmental factors in Jazan area. Survey data from 88 plant species related to 30 families were recorded in this study area, soil properties data from Forty stands (20 x 20 m each) were measured in the study area in April (2013-2014). Analysis data involved two steps: classification (using TWINSpan) and ordination (using CANOCO). The factors affecting the species distribution and correlation between the vegetation gradients and the edaphic variables are discussed. Eight major community types constitute the major part of the natural vegetation of the study area and are dominated by nine perennials: *Ziziphus spina-christi* (L.) Willd, *Calotropis procera* (Wild) R. Br., *Leptadenia pyrotechnical* (Forssk.) Decne., *Suaeda monoica* Forssk., *Panicum turgidum* Forssk., *Salvadora persica* L., *Acacia tortilis* (Forssk) Hayne, *Tamarix aphylla* (L.) Karst, and *Cyperus conglomerates* Rottb. The complex interaction of different environmental factors in relation to edaphic variables leads to variation of habitat types and different plant distribution communities. Data analysis showed that pH, moisture, electrical conductivity, organic carbon, calcium carbonate, bicarbonate, soil cations: sodium, potassium, calcium, and the sodium adsorption ratio are the main operating edaphic factors in Jazan area. The results of this study confirm that the study area is a subtropical desert and belongs floristically to the Sudanian territory and also that therophytes are the most frequent life-form in this region.

Key words: natural plant communities; plant distribution; soil properties; Jazan region.

INTRODUCTION

The aim of this paper is to examine the spatial pattern of floristic composition at species and community levels and to clarify the correlation between plant distribution and edaphic factors

of Jazan region, Saudi Arabia.

The southwestern region of Saudi Arabia is unique in regard to its nature, landform, climate and water availability [1]. Several ecological studies have been done in some of the phytogeographical regions of Saudi Arabia [2], [3], [4], [5], [6], [7], [8], [9], [10], [11], [12].

Most of these studies were based on qualitative field observations and interpretation of the authors. Although these studies were focusing on preparing floristic lists [2], and [10], [11], [12], some of them gave detailed description of the plant communities in relation to some ecological factors e.g. edaphic conditions, climate, aridity and topography [9], [13], [5], and [8] found that the complex interaction of different environmental factors in relation to altitude leads to variation of habitat types and different plant communities and vegetational belts.

[14] Identified Jazan region geomorphologically, by three main sectors; A) Mountains: E1-Sarwat mountains, B) Plains: 'Tihamah' coastal plains, and C) Islands: including those between Jazan city and Farasan islands.

Ecologically, [15], distinguished five ecosystems in Tihamah coastal plains namely: shoreline, sand formations, salt marshes, wadis and rocky hills. [16] discussed vegetation, species diversity and floristic relations in south-western Saudi Arabia and demonstrated remarkable change with altitude. There has been a broad discussion on the relationship among vegetation, soil factors and land use patterns [17], [18], [19], [20], [21], [22], [23], especially on the relationship between soil and vegetation distribution. The present study describes the structure of the plant communities and correlates their distribution with the edaphic variables of the habitat types. The primary questions were: is there any impact factor on plant distributions, Characteristics of natural plant communities with edaphic factors in Jazan region, Saudi Arabia.

MATERIALS AND METHODS

Study area

Jazan region located in the southwestern part of Saudi Arabia (lat. 16 ° 54' N & long. 42 ° 33' E). The research area is on coastal plains of Jazan region. The current study was performed in Jazan district, Saudi Arabia [Figure 1 A (Saudi Arabia) and B (The study area)]. The region is located in the southwestern part of the Kingdom of Saudi Arabia between the longitudes (42 - 43) east and latitudes (16 - 17) in the north, and bounded on the north and east Asir region to the west of the Red Sea along the coastline for about 330 km, and from the south and south-east as the Republic of Yemen, as has the average width of the area from east to west of about 100 km. The study period was for one year (April 2013 to April 2014). The present study focuses on using 40 stands (20 x 20 m each) [Fig. 2 A (Jazan-Jazan Dam), B (Sabya), C (Baysh), D (Abu Arish), and E (Samtah)], represented and covered the main physiographic variations of study region. Geologically, the study area belongs to the greater Afro-Arabian shield which is a part of the Precambrian crustal plate, generally exposed and locally covered by Tertiary volcanic rocks [24].

According to the climatic normal of Jazan region (averages of 2000-2010), Jazan area is generally hot during summer with the mean monthly air temperature ranges between 30°C during January and 40 °C during August. The maximum rainfall (18.0 mm) was recorded during December, while the minimum (0.8 mm) was attained during June. The climate diagram (Fig. 3) demonstrates dry period prevailing all the year round. The mean annual relative humidity of the study area is about 64.9%. The wind speed has a narrow range of variation (11- 12.2 km/h). Stormy wind with fine aeolian particles prevails during July-August with a northwestern direction [25].

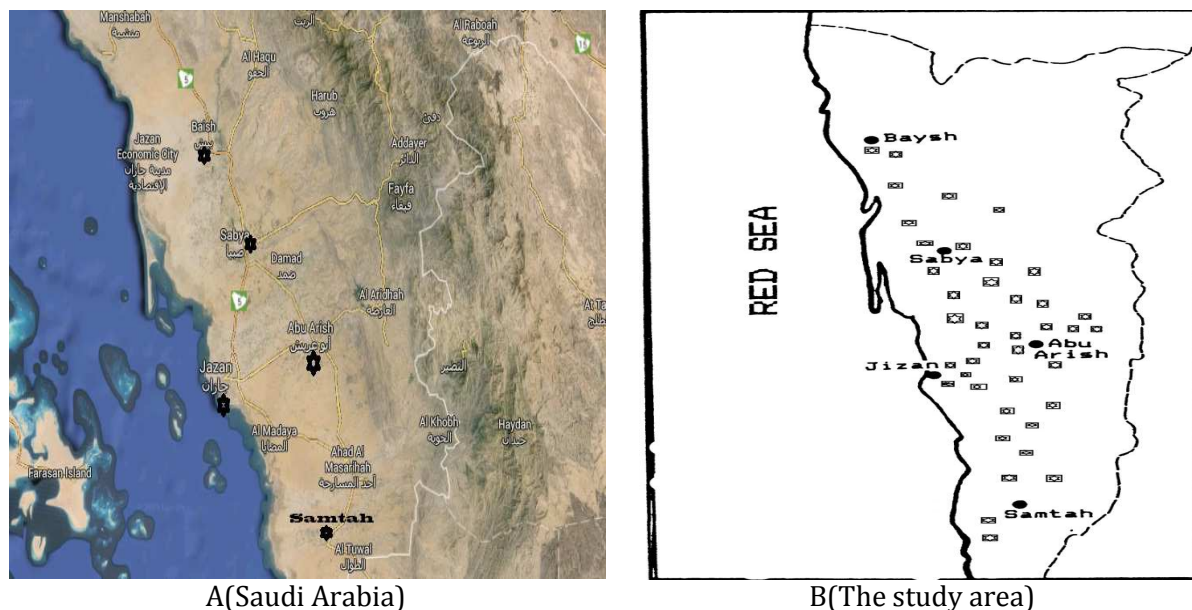


Fig. (1): showing (A) Saudi Arabia (B) showing location of the study area

Research design and sampling

Forty stands (20 x 20 m each) were selected and measured in the study area. In selecting each stand, care was taken to ensure reasonable degree of physiographic and physiognomic homogeneity of both habitat and vegetation. Sampling sites were selected, representing the most common habitats and herbaceous communities in the data as described by [26],[27],[28], and [29]. Soil samples measured and described by [26], [28], and [29]. A count-floristic list was carried out for each stand and relative frequency and density for each perennial species were calculated, while the relative cover was determined using the line intercept method. The sum of the relative frequency, relative density and relative cover gave the importance value (IV) for perennial species. The annual species were recorded in each stand. Composite soil samples taken from each stand to a depth of 50 cm. Soil samples were air-dried, crushed and sieved (< 2 mm) as described by [30], [31] and [32]. The Electrical conductivity (EC), pH and soluble ions were measured in soil saturation extracts. Na and K were measured by flame emission spectroscopy, Ca and Mg by atomic absorption spectroscopy, Cl and SO₄ by visible spectrophotometry, Organic matter (OM) was measured using a modified Walkley & Black method and total inorganic carbonates (TIC) was analyzed using a pressure calcimeter [33].



Fig.2: (A, B, C, D and E) showing plant diversity and sampling in location of the study area.

Analysis of plant community composition

The following three indices reflecting the characteristics of plant community were selected: Species richness: Margalef index (MA) was adopted, which is expressed as [34]: $MA = (S - 1) / \ln N$ (1) where S is the number of species, and N is the number of individuals of all species in a community. Species diversity: Shannon-Weaver index (H) was used, which can be calculated as [34]: $H = -\sum P_i \ln P_i$ (2) where $P_i = n_i / N$, n_i is the importance value of species i, and N is the sum of the importance value of all species in a community. Species ecological dominance: Simpson dominance index (D) was used, which is expressed as: $D = \sum (n_i / N)^2$ (3) where n_i is the number of species i in a community, and N is the number of all individuals in a community [34].

Vegetation and related environmental factors were analyzed using ordination techniques. A direct ordination method was used to enable us to test the environmental variables collected relevés. The analyses were conducted using CAP and Canoco program (Version 4.5) [35], [36]. Detrended Correspondence Analysis (DCA) was used to detect the length of the environmental gradient. Then, Canonical Correspondence Analysis (CCA) was applied because the data set was relatively heterogeneous and therefore, the length of ordination axes in DCA was relatively long [37]. Clustering methods were used to determine the patterns underlying species distribution and how plant communities may delineate habitat types in the study area.

Two Way Indicator Species Analysis [38] was used for this purpose. Floristic identification are according to [39],[40], [41], [42] and [43], [44], [45].

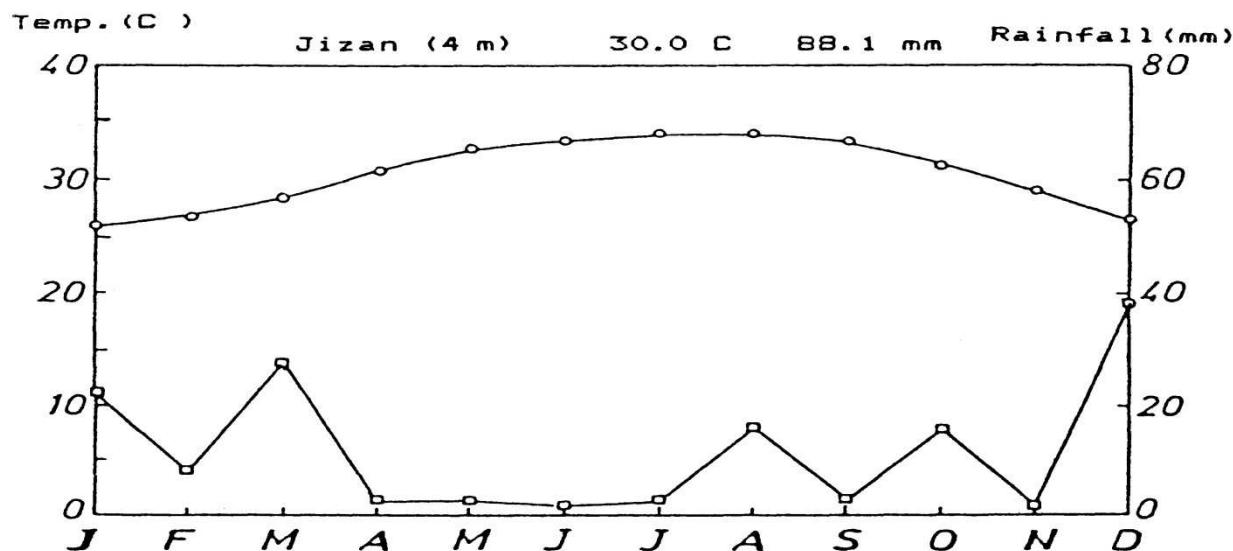


Fig.3: The climate diagram of the study area.

RESULTS

3.1. Analysis of plant community composition

Eighty eight plant species related to 30 families were recorded in the study area. They constituted about 46 perennial and 42 annual species: Gramineae 27.0%, Leguminosae 12.4%, Zygophyllaceae, Euphorbiaceae and Solanaceae together 17.9%, Asclepiadaceae, Amaranthaceae, Cappariaceae and Cruciferae make up 15.9%, while the other 22 families make up 26.8% of the total (Appendix). The most common perennials recorded were: *Tephrosia polifolia*, *Dipterygium glaucum*, *Panicum turgidum* and *Acacia tortilis* (presence percentage: 40.0-57.5%). *Salvadora persica*, *Ziziphus spina-christi*, *Leptadenia pyrotechnica*, *Calotropis procera*, *Suaeda monoica*, *Aerva javanica*, *Indigofera spinosa*, *Heliotropium arbainense*, and *Cassia italica* attain presence percentage ranging between 20.0 and 35.0%. The most common annuals in the study area are: *Cenchrus ciliaris*, *Chloris virgata*, *Tribulus terrestris*, *T. longipetalus*, *Euphorbia prostrata*, *Schouwia thebaica*, and *Trianthema portulacastrum*. Therophytes constituted about 48.3% of the total flora in the study area, followed by Chamaephytes (31.5%), Phanerophytes (10.1%), Cryptophytes (4.5%), Hemicryptophytes (5.6%). The majority of the recorded species belong to the sudanian and tropical territories.

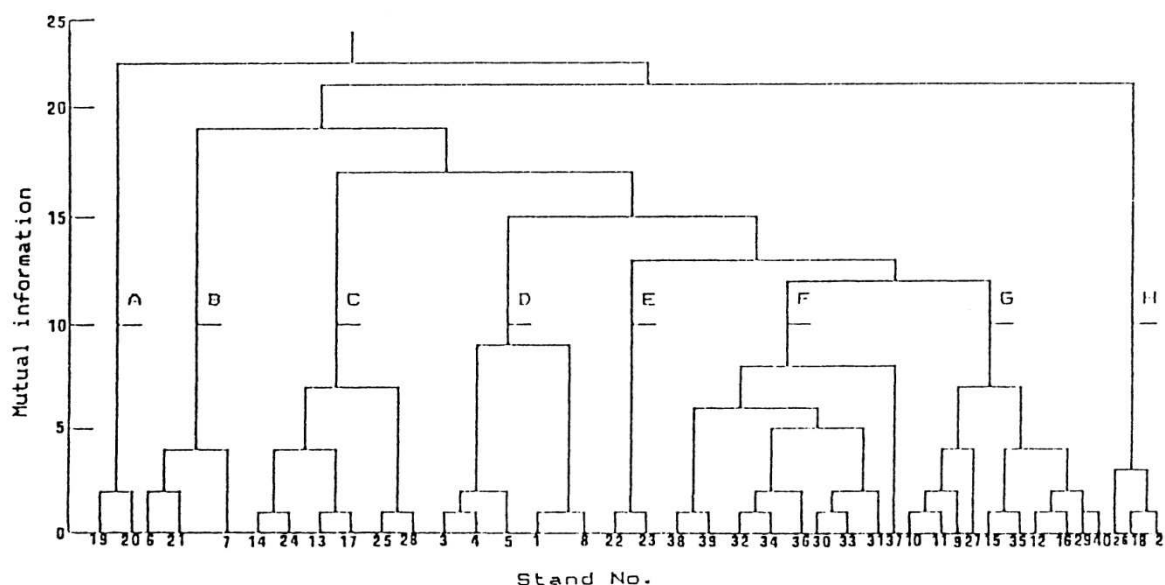


Fig.4:-The dendrogram resulting from the cluster analysis of stands representing the study area. The dashed line denotes the level at which the dendrogram yields eight distinct clusters:

- A: *Ziziphus spina-christi*
- B: *Calotropis procera*
- C: *Leptadenia pyrotechnica*
- D: *Suaeda monoica*
- E: *Panicum turgidum*
- F: *Salvadora persica*
- G: *Acacia tortilis*
- H: *Tamarix aphylla-Cyperus conglomeratus*.

3.2. Impact factors of association group distribution

The dendrogram resulting from the cluster analysis of stands representing the study area (Fig. 4 & Table 1), shows eight distinct clusters. Cluster 'A' comprises two stands dominated by *Ziziphus spina-christi* (IV=172.5/300), 'B' (3 stands) dominated by *Calotropis procera* (IV=181.7), 'C' (6 stands) dominated by *Leptadenia pyrotechnica* (IV=161.0), 'D' (5 stands) dominated by *Suaeda monoica* (IV= 159.9), 'E' (2 stands) dominated by *Panicum turgidum* (IV = 128.4), 'F' (9 stands) dominated by *Salvadora persica* (IV = 122.1), 'G' (10 stands) dominated by *Acacia tortilis* (IV = 86.2), 'H' (3 stands) and co-dominated by *Tamarix aphylla* and *Cyperus conglomeratus* (IV = 80 & 72.3). Shortly after rainfall, the soil surface of the study area is covered with dense growth of annual species e.g. *Zygophyllum simplex*, *Chloris virgata*, *nchrusciliaris*, *Tribulus terrestris*, *Tribulus longipetalus*, *Schouwia thebaica*, *Trianthem portulacastrum*, and *Boerhavia diffusa*.

Table 1. Means and standard deviation of the importance values (out of 300) of the perennial species in the different vegetational clusters resulting from the agglomerative cluster analysis of stands of the study area.

Plant species	Clusters							
	A	B	C	D	E	F	G	H
<i>Salvadora persica</i>	-	-	-	-	-	122.0 ± 38.0	5.2 ± 9.3	6.1 ± 8.6 6.1
<i>Acacia tortilis</i>	-	-	-	-	-	1.4 ± 1.4	86.1 ± 10.0	
<i>Panicum turgidum</i>	-	-	19 ± 29.4	-		2.1 ± 4.1		
<i>Dipterygium glaucum</i>	-	38.5 ± 10	25.6 ± 20.6	-		6.7 ± 17.8		

<i>Cassia italica</i>	84.3 ± 16.2	14.8 + 3.7	-	-		3.2± 7		
<i>Cassia senna</i>	40.4 ± 0.6		3.0 ± 6.7	-		-		
<i>Aeluropusmassuauensis</i>	-		-			24.9±47		
<i>Suaedamonoica</i>	-		-	158 - ± 60.4		2.3 ± 6.0		24.0 ± 2.6
<i>Leptadeniapyrotechnica</i>	-		161 ± 42.1			-	7.8+13.1	
<i>Euphorbia inarticulata</i>	-		-			24.7±35		
<i>Tephrosiapurpurea</i>	2.1 + 1.6	17.8 ± 2	69.2 ± 55.3			3±7.7		
<i>Arvapersica</i>	0.2 ± 0.3	8.7 ± 4.9	-			1.2±2.6		
<i>Cressacritica</i>	-		-	32.7 ± 9.4		-		14.8 + 4.5
<i>Sproboluspicatus</i>	-		-			11.2±21		
<i>Aeluropuslittoralis</i>	-		-			-		
<i>Sporobolusvirginicus</i>	-		-			-		
<i>Corchorusdepressus</i>	-		0.4 ± 0.8			10.5±28		
<i>Indigoferaspinosa</i>	-		1.8 ± 4.1			14±16.8		
<i>Tamarixaphylla</i>	172.5 ± 15.2		3.8 ± 8.6			-		
<i>Abutionpannosum</i>	-		0.3 ± 0.6			-		
<i>Calotropisprocera</i>	-	181.7 + 20	0.8 ± 1.7			-		
<i>Cyperusconglomeratus</i>	-		-			-		
<i>Aloe vera</i>	-		-			28.4±45		
<i>Arstolocheabracteolata</i>	-	17.6 ± 2.5	-			-		
<i>Ziziphus spina-christi</i>	-		-			-		
<i>heliotropiumbacciferum</i>	-		1.8 ± 4.0			0.9±2.5		
<i>Belipharisciliaris</i>	-		-			11.2±30		
<i>Fagoniabrugueri</i>	-		-			-		
<i>Citrulluscolocynthins</i>	-		-			-		
<i>Carallumapenicilla</i>	-		11.5 ± 25.8			-		
<i>Lyciumshawii</i>	-		1.8 ±			-		

			4.0					
<i>Cissusquadrangular</i>	-		-			-		
<i>Stippagrostispulmosa</i>	-		-			0.1±0.4		
<i>Panicumrepens</i>	-		-			-		

Table 2.Means and standard deviation of the different edaphic factors in the stands representing the different vegetational clusters of the study area. E.C: Electrical conductivity (?uS/cm), O.C.: Organic carbon (%), CaC03: Calcium carbonate (%), HC03: Bicarbonate (%), Na +,K+,Ca+ +, Mg+ +: Extractable cations (meq/1).

Clust ers	soil factors									
	moistu re (%)	pH	E.C. (µg/cm)	O.C. (%)	CaC03(%)	HC03(%)	Na+	K+	Ca++	Mg++
A	0.15 ± 0.05	8.50 ± 0.04	400 ± 50.0	1.5 + 0.1	0.7 ± 0.1	0.6 ± 0.2	4.5 ± 0.1	0.35 ± 0.1	9.65 ± 0.2	2.4 ± 0.3
B	1.8± 1.50	7.2 ± 0.51	138 ± 25.9	1.5 ± 0.57	3.0 ± 1.4	0.24± 0.05	2.3 ± 1.6	0.1 ± 0.1	28.8 ± 16.0	2.9 ± 0.29
C	0.67 ± 0.20	8.50 ± 0.34	165 ± 6.2	1.0 ± 0.33	2.2 ± 0.2	0.5 ± 0.02	3.2 ± 0.69	0.3 ± 0.1	7.25 ± 2.3	2.2 ± 0.7
D	4.2 ± 3.00	7.5 ± 0.72	469.0 ± 355.2	0.54 ± 0.74	3.2 ± 3.10	0.23 ± 0.11	44.7 ± 77.7	0.3 ± 0.38	266 ± 483	3.14 ± 0.6
E	1.5 ± 0.15	8.2 ± 0.20	110 ± 10.0	1.5 ± 0.2	1.5 ± 0.50	0.38 ± 0.01	2.4 ± 0.10	0.21 ± 0.01	3.0 ± 0.10	2.4 ± 0.2
F	1.75 ± 1.14	6.80 ± 0.36	563 ± 549	0.92 ± 0.5	0.78 ± 1.3	0.2 ± 0.1	368 ± 324	0.7 ± 2.6	459 ± 142	3.23 ± 0.29
G	0.85 ± 0.3	8.33 ± 0.6	322.5 ± 395	1.3a ± 0.8	3.6 ± 2.74	0.43 ± 0.16	5.0 ± 1.87	0.41 ± 0.27	29.7 ± 67.1	2.34 ± 0.8
H	1.5 2 ± 1.2	8.16 ± 0.94	495 ± 339.4	0.8 ± 0.50	1.7 ± 1.2	0.16 ± 0.05	6.12 ± 2.68	0.34 ± 0.07	92.3 ± 122	3.13 ± 0.73

3.3.Impact factors of vegetation indices

The results in (Table 2) demonstrate the variation in edaphic factors between the different groups of stands representing the eight clusters of the vegetation in the study area. It is clear that communities of *Salvadorapersica*, *Suaedamonoica* and *Tamarixaphylla-Cyperusconglomeratus* dominates saline soils with an E.C.ranging between 400 and 563 /µS/cm, while communities of *Panicumturgidum*, *Calotropisprocera* and *Leptadeniapyrotechnica* dominates sites with the lowest salinity levels (E.C: 110-165 ^uS/cm. The *Suaedamonoica* community type dominates the sites with high moisture content (4.2%), while the very dry sites are dominated by communities of *Ziziphus spina-christi*, *Leptadeniapyrotechnica* and *Acacia tortilis* (0.15-0.85%). All communities dominate ites with alkaline soil except *Salvadorapersica* dominates places with slightly acidic soil.*Suaedamonoica* dominates communities growing on soil with the lowest content of organic carbon (0.54%). The maximum level of calcium carbonate was recorded in the soil under communities of *Acacia tortilis*,*Suaedamonoica* and *Calotropisprocera* (3-3.55%). The soil bicarbonate content recorded its maximum level under *Ziziphus spina christi* and *LeptadeniaPyrotechnica* communities (0.51-0.6%), while the minimum level was recorded in soils co-dominated by *Tamarixaphylla-Cyperusconglomeratus*. With regard to soil soluble cations, soil dominated by *Salvadorapersica* attained the maximum levels (Na+: 368.3,K+: 3.7, Ca++: 458.9 & Mg++: 3.23 meq/1).The minimum levels of sodium and calcium was recorded in soil inhabited by

Panicumturgidum, while that of potassium was recorded in soil in habited by *Calotropisprocera*. The soil dominated by *Leptadeniapyrotechnica* showed the minimum level of magnesium. The ordination of stands indicated a clear distinction of the groups obtained by the agglomerative cluster analysis (Fig.5). Groups A and F lie nearly on the positive side of axis I and the negative side of axis II. Group C occupies the positivepart of both axes, while groups B and G occupy the positive sides of axis II and the negative side of axis I. In the meantime, groups D, E and H occupy the negative sides of both axes. Table 3 shows that soil pH, bicarbonate, sodium, and potassium calcium and sodium adsorption ratio have high significant correlation with axis I, while soil pH, moisture, electrical conductivity bicarbonate and calcium carbonate show significant correlations with axis II.

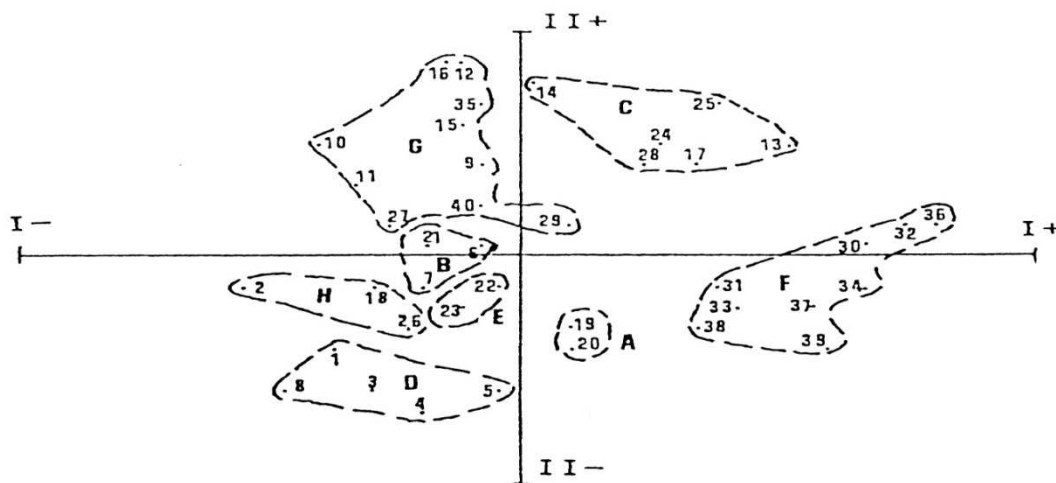


Fig. 5. Stand ordination of the study area on the first and second axes derived from the principal component analysis (PCA).The classification of stands by the cluster analysis is superimposed.

Table 3.Simple linear correlation coefficients (r) between the 1st & 2nd component axes as determined by the principal component analysis, and the edaphic variables (E.C: electrical conductivity, O.C.: Organic Carbon, *: P = 0.05, **: P = 0.01, ***:P =0.001).

Axe s	Soil variables											
	pH	Moi stur e	E.C .	CaC 03	O. C.	HCO 3	Na+	K+	Ca++	Mg ++	PAR	SA R
I	- 0.61 18* **	- 0.0 367	+0.03 67	- 0.3 302 **	- 0.15 10	- 0.42 68* **	+0.5252* **	+0.4951* **	+0.6279* **	+0.3087	+0.4968* **	+0.1023
II	+0.3594 *	- 0.3388 **	- 0.3649*	+3858* *	+1853	+3467*	- 0.2479	- 0.2226	- 0.2466	- 0.2856	- 0.2431	- 0.1385

DISCUSSION

Floristic Patterns with Ecological Gradients:

Impact Factors on Distribution, Characteristics of natural plant communities in Jazan region, Saudi Arabia demonstrate remarkable change from one site to other in the study area. The study area represents a continuation of the Sudanian tropical region with close similar climatic and topographic condition; therefore most of the recorded species in this study are afro-Tropical ones.

[46], regarded Southern Arabia and Ethiopia as a single phytogeographical unit, the Eritreo-Arabian province. In the meantime, [47], reported that the floristic composition of Southwest Arabia, especially those of the interior deserts show some floristic affinities to the Irano - Turanian and Mediterranean territories.

According to Koppen's climatic system, the Tihamah coastal area lies within the subtropical dry zone of the desert, characterized with high temperature ranges (29-38 °C), relative humidity(64.2 %), and low rainfall (0.8-19.0 mm). Summer rainfall is connected with the influence of Indian monsoon, while, the winter rainfall is related to the Mediterranean regime [48]and [42].Since the local climate over the coastal plains of Tihamah is almost uniform, therefore, the distribution and species composition of the plant communities seem to be more affected by various edaphic factors.Shortly after rainfall, the soil surface of the study area is covered with dense growth of annual species,e.g. *Zygophyllum simplex*, *Chloris virgata*,*Cenchrusciliaris*, *Tribulusterrestris*, *T. longlpetalus*, *Schouwiathebaica*, *Trianthemaportulacastrum*, and *Boerhaviadiffusa*. The amount and seasonally of rainfall together with the high temperature controls the growth of these communities.

[49], reported similar communities inthe coastalplain, foothills and wadis of the gulf of Elat and demonstrated that these communitiestolerate lower annual temperature (23-26C) and receivehigher annual rainfall (5-30 mm) ascompared to habitat condition in Tihamah plains(29-38 °C& 0.8-19 mm respectively). [50], reported that *Ziziphus spina-christi*growsin diffusedpattern on the Mediterranean coastalplains and in the Jordan Valley, and that *Tamarixaphylla* has a very wide range of distribution inNorth & South Africa and Western Asia, anddescribed*Suaedamonoica* as thermophilic species which cannot withstand even mild winter.[51], reported *Acacia tortills* as a Sudanian treedominating the more arid places of the southern coast of Sinai. [52],and [13].Described similar communitiesalong the coastal plains of the Saudi Red Seacoast.[42], gavedetaileddescription of the habitats and plantcommunities in Tihamahplains on the coast ofYemen. Kassas and [53], and [54],described similar communitiesalong the Egyptian Red Sea coast.

According to [43],the southern Arabian phytogeographicalregion(including Yemen) contains 2000-2500species of the flowering plants[55]. The most important tropical genera in Arabia are *Acacia*, *Indigofera*, *Euphorbia*, *Solanum*, *Aristida*, *Caralluma*, *Tephrosia*, *Pulicaria*, *Barleria*, *Grewia*, *Commiphora*, and *Cadaba*. About 20% of the floristic elements of southern Arabia are endemic [56],[57].

The present study records 88 species in 30 familiesinthe Tihamah coastal plains of Jazan region, of which 46 are perennial and 42 annual. Multivariate analysis including classification and ordination can provide more detailed and comprehensive information on the patterns in vegetation and the response of plant species to the underlying gradients [58], [59].The classification of the stands representing the study area using the agglomerative cluster analysis led to the identification of eight major plant community types forming the major part of the natural vegetation. These communities are dominated by nine perennials: *Ziziphusspina Christi* growing mainly inwadis characterized with fine soil, with high water retaining power. While communities dominated by *Calotropisprocera* always inhabit sand plains around villages and farms. *Leptadeniapyrotechnica*, *Panicumturgidum*, and *Acacia tortilis* community types dominate the inland hyperarid sand plains forming a poor savanna scrub. Communitiesdominatedby*SalvadoraPersica*occupy old wadi beds characterized with its salt

affected sandy soil. This habitat represents a transition between sand plains and salt marshes. In the meantime, communities dominated by *Suaedamonoica*, *Tamarixaphylla* and *Cyperusconalomeratus* occupy the salt marsh habitat type.

The study area represents a continuation of the Sudan region which has close similar climatic and topographic features, therefore most of the recorded species are Afro-Tropical ones, and belong to the Paleotropic kingdom [76].

Scrub savanna include a wide range of vegetation types and are defined in many different ways. [61], grouped savanna in three groups: climatic, edaphic and anthropogenic savanna. Synthetically, savanna can be characterized by a continuous herbaceous layer and a discontinuous cover of shrubs and trees [62].

Besides spatial variations in the species composition of plant communities, the composition of life-forms reflects the response of vegetation to variation in certain environmental factor. The life-form spectra is thought to be either a hereditary adjustment to environment [63], or represent the residual effects of some historical, climatic or biotic conditions on the population of the plant [50]. In this study, therophytes are the most frequent life-form in the study area, followed by chamaephytes, phanerophytes, cryptophytes, hemicryptophytes.

The dominance of both therophytes and chamaephytes over the other life-forms in Tihamah coastal plains seem to be a response to the hot dry climate, topographic variations, and biotic influence. A comparison of the life-form spectra, indicates that the Southern region contains more therophytes, chamaephytes and less hemicryptophytes (48.3, 31.5 & 5.6%) than the Eastern region (46, 23 and 6%) respectively. In the meantime, it contains almost the same figure of cryptophytes about 4.5% and about half of the phanerophytes (10.1%).

The correlation analysis demonstrated that pH, bicarbonate, sodium, potassium, and calcium to sodium adsorption ratio have high significant correlation, while soil pH, moisture, electrical conductivity, bicarbonate and calcium carbonate showed significant correlations as (Fig. 5). The ratio $\text{Na}^+ / (\text{Ca}^{++} + \text{Mg}^{++})$, and $\text{K} \% / (\text{Ca} \% + \text{Mg} \%)$ determines the adsorption pattern of these cations by the exchange complex [50], [63], [11], [12]. However, our results showed that silt and clay content, organic matter and electrical conductivity are mainly related to vegetation distribution. In addition, [64] reported that edaphic factors provide the primary explanation for describing the distribution of plant communities in the Western Desert of Egypt. Soil texture controls dynamics of soil organic matter in many simulation models or organic matter decomposition and formation [65], [66], [67], and influences infiltration and moisture relation and the availability of water and nutrients to plants [68]. Under the condition of low and irregular rainfall which prevail in the study area, local topography and water redistribution within the local landscape [69], [70].

In accordance with the other studies of the desert ecosystems [71], [5], [7], [72], [73], [74], [60], the present study indicated that the distribution of the recognized plant communities in Jazan region is affected mainly by physiographic features, climatic conditions as well as soil attributes, and that these communities are modified both in their distribution and abundance by the effects of some other factors that threaten the plant life in Tihamah area including:- a) Agricultural activities, b) Heavy grazing, c) Woodfuel cutting and d) Termites [13], [76], [77].

Spatial distribution of plant species and communities over a small geographic area in desert ecosystems is related to heterogeneous topography and landform pattern [78]. The heterogeneity of local topography, edaphic factors, microclimatic conditions lead to the variation in the distributional behavior of the plant associations of the study area.

On the tropical scale, [79], detected some similar communities on an arid misty mountain plateau in Sudan and concluded that the species composition, at all spatial scales is directly or indirectly related to variation in temperature and moisture. [80] While dealing with the vegetation types and patterns in Senegal described communities of *Calotropis procera*, *Acacia tortilis* and *Ziziphus spp.* in the Sahelian grassland.

In conclusion, this study showed that the vegetation in the study area belongs to the Sudanian territory. Edaphic factors play the leading role in the distribution of the different plant community types. Human and animal impacts modified the distribution and abundance of the plant species and life-forms spectrum.

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Appendix

Floristic List of the Recorded Species in Coastal Plains of Jazanregion, Saudi Arabia.

1. Acanthaceae: *Blepharisciliaris* (CL.) B.L.Burt
2. Aizoaceae: *Aizooncanariense* L.; *Trichanthemaporulacarrum* L.
3. Amaranthaceae: *Aerobajavanica* (Burro. f.) Juss.; *Amaranthusgraecizans* L.; *Digeramuricata* (L.) Mart.
4. Aristolochiaceae: *Aristolochiabracteolata* Lain.
5. Asclepidaceae: *Calotropisprocra* (L.) Aitf.; *Carallumapenicillata* (Defl.) N.E. Br.; *Leptadeniapyrotechnica* (Forssk.) Decne.
6. Boraginaceae: *Heliotropiumarabianense* Fresen.
7. Capparaceae: *Capparis decidua* (Forssk.) Edgew.; *Capparis spinosa* L.; *Cleome viscosa* L.; *Dipterygium glaucum* Decne.; *Crynandropsis gynandra* (L.) Briq.
8. Chenopodiaceae: *Halopeplisperfoliata* (Forssk.) Bunge ex Schweinf.; *Suaeda monoica* Forssk.
9. Commelinaceae: *Aneilematacazeanum* Hochst. ex. C.B.C1.
10. Compositae: *Launaea* sp.; *Xanthium spinosum* L.
11. Convolvulaceae: *Cressacrerica* L.
12. Cruciferae: *Anastaticahierochuntica* L.; *Savignya parviflora* (Del.) Webb *Schouwiatehaica* Webb
13. Cucurbitaceae: *Citrullus colocythis* (L.) Schrad.
14. Cyperaceae: *Cyperus conglomeratus* Rottb.
15. Euphorbiaceae: *Euphorbia hirta* L.; *Euphorbia articulata* L.; *Euphorbia prostrata* Ait.; *Euphorbia aculeata* L.; *Jatropha glauca* Vahl
16. Gramineae: *Aeluropus lagopoides* (L.) Trill. ex Thw.; *Aeluropus littoralis* (Gouan) Parl.; *Cenchrus ciliaris* L.; *Chloris virgata* Sw.; *Crypsis aculeata* (L.) Alton; *Cutandia cotoma* (Forssk.) Trabut; *Cutandia emphitica* (Spreng.) Benth.; *Dacryloctenium aegyptium* (L.)
- P. Beauv.; *Digitaria faciliaris* (Retz.) Koel; *Digitaria sanguinalis* (L.) Scop.; *Echinochloa colona* (L.) Link; *Eragrostis ciliaris* (All.) Vign.-Lut.; *Imperata cylindrica* (L.) Raeusch; *Panicum repens* L.; *Panicum turgidum* Forssk.; *Phalaris minor* Retz.; *Polypogon monspeliensis* (L.) P. Desf.; *Rosieria cristata* (L.) Tzvelev.; *Setaria verticillata* (L.) P. Beauv.; *Setaria viridis* (L.) P. Beauv.; *Sorghum bicolor* (L.) Moench; *Sporobolus plumosa* (L.) Munro ex T. Anders.; *Sporobolus spicatus* (Vahl) Kunth; *Sporobolus virginicus* (L.) Kunth
17. Leguminosae: *Acacia ehrenbergiana* Hayne; *Acacia mellifera* (Vahl) Benth.; *Acacia raddiana* Savi; *Acacia torrilis* subsp. *radiana* (Forssk.) Hayne; *Indigofera hochstetteri* Baker; *Indigofera spinosa* Forssk.; *Rhynchosia minima* (L.) DC. var. *memnonia* (Del.) Cooke.; *Senna alexandrina* Forssk.; *Senna italica* Mill.; *Tephrosia apollinea* (Del.) Link; *Trigonella hierosolymitana* Boiss.
18. Liliaceae: *Aloe vera* L
19. Malvaceae: *Abutilon pannosum* (Forst. f.) Schlecht.
20. Portulacaceae: *Portulaca oleracea* L.
21. Rhamnaceae: *Ziziphus spina-christi* (L.) WiUd.
22. Salvadoraceae: *Salvadora persica* L.
23. Scrophulariaceae: *Strigahermomhica* Benth.
24. Solanaceae: *Datura innoxia* Mill.; *Datura stramonium* L.; *Lycium shawii* Roem. & Schult.; *Solanum incanum* L.; *Solanum nigrum* L.
25. Tamaricaceae: *Tamarix mannifera* (Ehrenb.) Bunge
26. Tiliaceae: *Corchorus depressus* (L.) Stocks

27. Urticaceae: *Urticaurens*L.
28. Vitaceae: *Cissusquadrangularis*L.
29. Zygophyllaceae: *Fagoniabrugueri*DC.; *Fagoniaindica*Burro. f.;
*Tribuluspentandrus*Forsk.; *Tribulusterrestris*L.; *Zygophyllum album* L.;
*Zygophyllumsimplex*L.
30. Cleomaceae *Cleome viscosa*L.