



Research Paper

PHYTOCHEMICAL ANALYSIS OF MARINE MACROALGA *Caulerpa racemosa* (J. AGARDH) (CHLOROPHYTA - CAULERPALES) FROM TIRUNELVELI DISTRICT, TAMILNADU, INDIA

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Abstract

Phytochemical screening providing important ideas for the development of new drugs against deadly diseases. Recently, a number of studies have been reported on the phytochemistry of marine algae across the world. The marine macroalga *Caulerpa racemosa* methanol solvent extract was prepared by cold percolation method. The shade dried seaweed powder and its crude extract were tested for organoleptic properties, fluorescence analysis, and qualitative phytochemical analysis for the identification of active bio-molecules. Methanol solvent extract of *Caulerpa racemosa* showed the presence of a number of metabolites such as phenolic groups, saponins, tannin, flavonoids, carbohydrates, reducing sugars, Xanthoproteins. This suggest that the *Caulerpa racemosa* seaweed can be used as antimicrobial (antiviral, antifungal and antibacterial), antiparasitic, anti-inflammatory, antifeedent, antioxidant, antiallergenic, antithrombic and antiulcer agents in the near future.

Key words: Seaweed, Phytochemistry and Secondary metabolites.

INTRODUCTION

Seaweeds are wonder plants of the sea and considered as medicinal food of the 21st century. Marine macroalgae, popularly known as seaweeds are potential renewable resources in the marine environment. About 6000 species of seaweeds have been identified and are grouped into green (Chlorophytes), brown (Phaeophytes) and red (Rhodophytes) algae. Marine algae from Indian coasts amounting to 844 species (including forma and varieties) are distributed among 217 genera. Seaweeds are primitive non-flowering plants without true root, stem and leaves. They grow in the intertidal, shallow and deep sea areas up to 180 meter depth and also in estuaries, backwaters and lagoons on solid substrates such as rocks, dead corals, pebbles, shells, mangroves and other plant materials (Anatharaman, *et al.*, 2007; Sakthivel, 2007).

Marine organisms are source material for structurally unique natural products with pharmacological and biological activities (Faulkner, 2001). Among marine organisms, the macroalgae occupy an important place as a source of biomedical compounds (Selvin and Lipton, 2004; Manilal *et al.*, 2009^{ab} and Manilal *et al.*, 2010). About 2400 natural products have been

isolated from macroalgae belonging to the classes Rhodophyceae, Phaeophyceae and Chlorophyceae (Faulkner, 2001).

Seaweeds known as medicinal are rich in secondary metabolites which include alkaloids, glycosides, flavonoids, saponins, tannins, steroids, related active metabolites, which are of great medicinal value and have been extensively used in the drug and pharmaceutical industry (Eluvakkal *et al.*, 2010). Recently, their value as a source of novel bioactive substances has grown rapidly and researchers have revealed that marine algal originated compounds exhibit various biological activities (Kim and Wijesekara, 2010; Wijesekara and Kim, 2010; Wijesekara *et al.*, 2010 and Wijesekara *et al.*, 2011).

In recent years, the seaweeds serve as an important source of bioactive natural substances (Plaza, *et al.*, 2008 and Smit, *et al.*, 2004). Moreover, many metabolites, which isolated from marine algae, have shown to possess bioactive effects (Faulkner, 2002 and Kim *et al.*, 2005). Therefore, recently a new trend has been arisen to isolate novel bioactive compounds and constituents from edible seaweeds. (Li, *et al.*, 2011). Phytochemical analysis of seaweeds can help the manufacturers for identification and selection of raw materials for drug production.

Seaweeds are the economic potential resource in marine environment and it will be a great significance in drug development (Fuesetani, 2000). Seaweeds known as medicinal are rich in secondary metabolites which include alkaloids, glycosides, flavonoids, saponins, tannins, steroids, related active metabolites, which are of great medicinal value and have been extensively used in the drug and pharmaceutical industry (Eluvakkal *et al.*, 2010).

A greater diversity in biochemical composition of seaweeds paves way to explore variety of compounds in their bodily composition with a wide range of physiological and biochemical characteristics, many of which are rare or absent in other taxonomic groups (Holdt and Kraan, 2011). The secondary metabolites of seaweeds such as Isoprenoids (terpenes, carotenoids, steroids), polyketides, phlorotannins, amino-acid derived natural products (alkaloids), and shikimates (flavonoids) have always attracted the interest of biochemists because of their diversity as compared with those present in the leaves of higher plants (Manilal *et al.* (2009).

Seaweeds were rich in dietary fiber (>50% dry weight), particularly in the soluble form (Mabeau *et al.*, 1993); Fleury *et al.* (1991) reported that the physico-chemical properties of seaweed powder could be assumed to reflect those of the present fibre. Moreover, since seaweed proteins are closely related to the cell wall polysaccharides (Fleurence, 1999), they may also play a role in the physicochemical properties, such as water holding.

Phytochemical screening providing important ideas for the development of new drugs against deadly diseases. Recently, a number of studies have been reported on the phytochemistry of plants across the world (Edeoga, *et al.*, 2005; Aliyu, *et al.*, 2008; Ayoola, *et al.*, 2008; Johnson, *et al.*, 2008; Maridass, *et al.*, 2008a; Maridass, *et al.*, 2008b; Ujowundu, *et al.*, 2010; Rafia Rasool, *et al.*, 2010; Usha and Bopaiah, 2011 and Hamad, *et al.*, 2011), in particular seaweeds (Poonam, 2011; Ganga Rao *et al.*, 2011; Jayasree *et al.*, 2012; Venkatesh *et al.*, 2011; Johnson, *et al.* 2012, and Rajasulochana, *et al.*, 2009) were reported.

Saponins are considered as a key ingredient in traditional Chinese medicine and are responsible for most of the observed biological effects. Saponins are known to produce inhibitory effect on inflammation. Flavonoids in human diet may reduce the risk of various cancers, as well as preventing menopausal symptoms. Its potent water soluble antioxidants and free radical scavengers prevent oxidative cell damage and have strong anti-cancer activity. They show anti-allergic, anti-inflammatory, anti- microbial and anti-cancer activity (Cushnie and Lamb 2005; De Sousa *et al.*, 2007 and Yadav *et al.*, 2011). Phenolic phytochemicals have antioxidative, antidiabetic, anticarcinogenic, antimicrobial, antiallergic, antimutagenic and anti-inflammatory properties (Arts *et al.*, 2005 and Scalbert *et al.*, 2005).

The secondary metabolites of seaweeds have always attracted the interest of biochemists because of their diversity as compared with those present in the leaves of higher plants. Isoprenoids (terpenes, carotenoids, steroids), polyketides (Phlorotannins), amino-acid-derived natural products (alkaloids), and shikimates (flavonoids) are the major groups of

secondary metabolites found in algae (Mendis and Kim, 2011). In this background, the present study intended to evaluate the qualitative and phytochemical (secondary metabolites) analysis in the seaweed *Caulerpa racemosa* collected from the Tirunelveli District Tamil Nadu, India was studied.

MATERIALS AND METHODS

Marine Alga *Caulerpa racemosa* (J. Agardh)

Caulerpa racemosa Thalli are pale green 5-6 cm tall. Plants in larger or smaller group possess rhizome with creeping, cylindrical and highly branched. Rhizoids arise from the lower parts of prostrate rhizome. Crowded with bud-like branchlets in spherical shape, sometimes oval not constricted at the base. They are found growing on sandy, rocky or silt-covered substrate attached by numerous rhizoids arising from the creeping stolon. It was observed that the thalli were rarely associated with other macroalgae (Iyengar, (1933) and Umamaheswaran Rao, 1987^{ab}). *Caulerpa racemosa* distributed in Maharashtra (Bombay, Malwan), Gujarat (Dwarka, Okha, Saurashtra, Veraval), Tamil Nadu (Tirunelveli, Idinthkarai; Kuttapuli, Palk Bay, Pamban, Mandapam, Tuticorin, Krusadai Island,(Gulf of Mannar), Kerala(Kovalam), Andhara Pradesh(Visakapatnam), Goa, Karnataka, Lakshadweep Island and Andaman and Nicobar Islands.

Collection of *Caulerpa racemosa* (J. Agardh)

The macro alga *Caulerpa racemosa* were collected from, Kuttapuli Tirunelveli District, Tamil Nadu, immediately after collection, the macro alga washed in sea water and the epiphytes, associated organisms and other extraneous matter were removed. After subsequent washing in fresh water the alga were shade dried for two weeks continuously. The shade-dried seaweed was partially powdered using domestic blender and stored in air tight container for further experiments.

Preparation of Extracts

From these stock, secondary metabolites of seaweed (100g), was extracted successively using (150mL) solvent methanol the sample were kept in dark for 96 hour. After incubation, the extract thus obtained was decanted and filtered. The clear extract was subsequently concentrated using rotary vacuum evaporator and kept in dark bottles in 4° C until use (Johnson *et al.*, 2012.)

Physico-Chemical parameters of seaweed

The *C. racemosa* powder was used for physico-chemical, fluorescent and phyto-chemical analyses. The procedures recommended in Indian Pharmacopoeia (Anonymous, 1966; 1985; 1996) were followed.

Fluorescence Analysis

Fluorescence characteristics of the seaweed powder as such and after treating them with chemical reagents were observed in day light. Fluorescent analyses of the seaweed powder were carried out according to the methods of Chase and Pratt (1949) and Kokoshi *et al.*, (1958) Kokate (2005); Nazish *et al.*(2009); Arun Kumar *et al.*.(2011).

Behavior of seaweed powder was treated with different reagents (NaOH, conc. HNO₃, con. HCl, con. H₂SO₄, 5% Acetic acid, 5% FeCl₃, ammonia, picric acid) and colour change was observed in natural light (Kay, 1938; Johansen ,1940; Pratt and Chase, 1949 and Nazish *et al.*, 2011).

Organoleptic properties

The organoleptic characteristics of alga powered samples namely their appearance and colour in day light, smell and their taste were also studied. Organoleptic evaluation seaweed powder were carried out according to the methods of Kokate *et al.*, 2005; Arun Kumar and Paridhavi, 2011).

Qualitative phytochemical Analysis

The different qualitative chemical tests were performed for establishing profile of algal extract for its chemical composition. Qualitative phytochemical analyses were done using the procedures of Kokate *et al.*, (1995) Sofowara (1993), Trease and Evans (1989),Harborne (1973), Brindha,(1991), Edeoga *et al.*, (2005), and Savithamma *et al.*, (2011) .

Swelling capacity (SWC) of seaweed *C. racemosa* was analyzed by the bed volume technique after equilibrating in excess solvent (Kuniak and Marchessault, 1972). Water holding capacity (WHC) of seaweed *C. racemosa* was measured by the modified centrifugation method described by Suzuki *et al.*, (1996). The WHC of seaweed was expressed as the weight of grams of water held by 1 g of sample (dry weight).

RESULTS AND DISCUSSION

The Organoleptic characteristics of alga powered sample results revealed that the colour is green, aromatic smell, Tasteless and soft nature when fresh (Table.1) The characteristic fluorescent properties or colours emitted by the powdered alga *C. racemosa* before and after treating with various reagents were recorded. The powdered alga as such appeared green under daylight and Ordinary light.

After treating with various reagents, under daylight, it showed different shades of green and brown. However, ordinary light, acidic and alkaline solutions of concentrated HNO₃, HCL, H₂SO₄, 50% H₂SO₄, 1N HCL, 50% HNO₃, 5% KOH, MeOH, 1N NaOH, 5% acetic acid, 5% FeCl₃, Ammonia and Picric acid, with light green, intense green and dark green colours respectively (Table 2& 3). The characteristic fluorescent properties or colours recorded through this study could be used as a standard in the identification and authentication of the alga *C. racemosa* in its crude form. The ash value indicates the presence of inorganic ions. During ash preparation organic matter gets oxidized and certain amount of volatile elements are lost. (Trease, 1961).

Preliminary phytochemical screening of twenty four different chemical compounds (steroids, alkaloids, phenolic groups, saponins, tannins, flavonoids, anthraquinones, reducing sugars, triterpenoids, terpenoids, cardiac glycosides, glycosides, phlobatannins, quinones, aromatic acids, essential oils, anthocyanins, leucoanthocyanins, Emodins, gum and mucilage, carbohydrates, Coumarins, aminoacids and xanthoprotein) were tested in methanolic extract. The results showed that the presence of steroids, phenolic groups, saponins, tannin, flavonoids, terpenoids carbohydrates, reducing sugars, anthraquinones, Emodins coumarins, and xanthoproteins. Alkaloids, triterpenoids, glycosides, cardiac glycosides, aminoacids, quinnone, gum and mucilage, anthocyanins, leucoanthocyanins, aromatic acids, did not show any positive result for their presence in any of the extract tested. Methanol extract showed the presence of maximum number (10) of compounds. (Table 4).

Table . 1. Organoleptic properties of the alga *C. racemosa*

S. No	Character	When fresh	After drying 20 days	Powder
1	Colour	Green	Light green	Light greenish brown
2	Odor	Aromatic	Aromatic	Aromatic
3	Taste	Tasteless	Tasteless	Sour
4	Texture	Soft	Crispy	Soft

Table 2. Fluorescence analysis of *C. racemosa* powder in different chemical reagents in ordinary light

S. No	Particulars of treatment	Under ordinary light
1	Powder as such	Green
2	Powder + 50% H ₂ SO ₄	Dark Intense green
3	Powder + 1N HCL	Grey green
4	Powder + 50% HNO ₃	Light grey
5	Powder + 5% KOH	Light green
6	Powder + MeOH	Light green
7	Powder + 1N NaOH	Light green

Table 3. Fluorescence analysis of *C.racemosa* powder in different chemical reagents in natural light

S. No	Drug treatment	Seaweed Powder
1	Powder as such	Green
2	Powder + 1N NaOH	Grey colour
3	Powder + con. HNO ₃	Yellowish colour
4	Powder + con. HCL	Intense light green
5	Powder + con. H ₂ SO ₄	Intense black colour
6	Powder + 5% acetic acid	Light green
7	Powder + 5% FeCl ₃	Light brown
8	Powder + Ammonia	Light green
9	Powder + Picric acid	Yellowish green

Table 4. Qualitative determination of Phytochemicals of alga *C.racemosa*

S. No	Phytochemicals	Inference
1	Alkaloids	-
2	Steroids	-
3	Reducing sugar	++
4	Tannins	++
5	Pholobatanins	+
6	Saponins	+
7	Coumerins	+
8	Flavonoids	+++
9	Terpenoids	-
10	Triterpenoids	-
11	Cardiac Glycosides	-
12	Glycosides	-
13	Anthraquinones	-
14	Phenolic compounds	++
15	Quinones	-

16	Aminoacids	-
17	Essential oil	-
18	Aromatic acid	-
19	Xanthoprotein	+
20	Carbohydrates	+
21	Anthocyanins	-
22	Leucoanthocyanins	-
23	Emodins	-
24	Gum and mucilage	-

(+++ very good);(++ good);(+) Positive; (-) Negative

In the present study, presence of steroids, phenolic groups, saponins, tannin, flavonoids, carbohydrates, coumarins, and Xanthoproteins. The present study was qualitative analysis of phytochemicals of crude extract was directly coincided with (Rajasulochana, *et al.*, 2009; Hebsibah *et al.*, 2011 ; Ganga rao *et al.*, 2011; Venkatesh, *et al.*, 2011; Poonam, 2011; Jeeva *et al.*,2012; Jayasree *et al.*, 2012 and Johnson, *et al.*, 2012) reports. These observations cited on phytochemical compounds support our findings on the usefulness of seaweeds in traditional medicament and added a note on the phytochemical history.

Tannins are known to possess general antimicrobial and antioxidant activities (Rievère *et al.*, 2009). The present study confirms the presence of tannin in methanol extract of *C. racemosa*. It suggests that the methanol extract of *C. racemosa* with tannins, can be used as antimicrobial agents. Tannins have been found to have antiviral, antibacterial, antiparasitic effects, antiinflammatory, antiulcer and antioxidant property for possible therapeutic applications (Kolodziej and Kiderlen, 2005).

Saponins are considered as a key ingredient in traditional Chinese medicine and are responsible for most of the observed biological effects. Saponin possesses specific physical, chemical and biological activities that make them useful as drugs. In medicine, it is used in hyper cholestrolaemia, hyperglycemia, antioxidant, anticancer, anti inflammatory and weight loss, *etc* (Jeeva *et al.*, 2012). Saponins have been implicated as bioactive antibacterial agents of plants (Mandal, *et al.*, 2005 and Manjunatha, 2006). Some of these biological properties include antimicrobial, anti-inflammatory, antifungal, antifeedent, and hemolytic effects (George *et al.*,2002; Xu *et al.*, 1996; De-Lucca *et al.*,2005 and Mohanta *et al.*, 2007).

The plant extracts were also revealed to contain saponins which are known to produce inhibitory effect on inflammation (Just,1998). Saponins have the property of precipitating and coagulating red blood cells. Some of the characteristics of saponins include formation of foams in aqueous solutions, hemolytic activity, cholesterol binding properties and bitterness (Sodipo, 2000 and Okwu, 2004). The present study confirms the presence of Saponin in methanol extract of *C. racemosa*. It suggests that the methanol extract of *C. racemosa* with saponins, can be used as antimicrobial, anti-inflammatory and antifungal agents.

The present study confirms the presence of steroid in extract of *C. racemosa*. Steroids have been reported to have antibacterial properties (Raquel, 2007) and they are very important compounds especially due to their relationship with compounds such as sex hormones (Okwu,

2001). It suggests that the extract of *C. racemosa*, can be used as antimicrobial agents. Coumarin has been used as anti-coagulant drugs and to treat lymphedema.

Flavonoids are hydroxylated phenolic substances known to be synthesized by plants in response to microbial infection and they have been found to be antimicrobial substances against wide array of microorganisms *in vitro*. Flavonoids, the major group of phenolic compounds reported for their antimicrobial and antiviral activity. The presence of Flavonoids in the methanol extract of *C. racemosa*. It suggests that the methanol extract of *C. racemosa* with flavonoids, can be used as antioxidant, antimicrobial, anti-inflammatory, antifungal and anti-cancer agents in the pharmaceutical industry. Flavonoids in human diet may reduce the risk of various cancers, as well as preventing menopausal symptoms. Its potent water soluble antioxidants and free radical scavengers, which prevent oxidative cell damage and have strong anti-cancer activity. They show anti-allergic, anti-inflammatory, anti-microbial and anti-cancer activity (Cushnie and Lamb 2005 ; De Sousa *et al.*, 2007 and Yadav *et al.*, 2011).

Seaweed extracts contain rich source of phenolic compounds (Athukorala *et al.*, 2003; Heo *et al.*, 2005). In the present study showed the presence of phenolics was confirmed by the qualitative analysis in the crude extract of the seaweed. *C. racemosa*. Phenolic compounds are commonly found in plants, including seaweeds, and have been reported to have a wide range of biological activities including antioxidant properties (Duan *et al.*, 2006; Kuda *et al.*, 2007; Wang *et al.*, 2009; Athukorala *et al.*, 2006), therefore the seaweed extracts could have potential applications in food industries (Yan *et al.*, 1996). The phenolic compounds are one of the largest and most ubiquitous groups of plant metabolites (Singh *et al.*, 2007). They possess biological properties such as anti-apoptosis, anti-aging, anti-carcinogen, anti-inflammation, anti-atherosclerosis, cardiovascular protection and improvement of endothelial function, as well as inhibition of angiogenesis and cell proliferation activities (Han *et al.*, 2007 and Yadav and Agarwala, 2011).

Capacity increase in the water holding of the *C. racemosa* alga powder with temperature. Such increase was probably related to the increase in the solubility of fibres and proteins (Fleury *et al.*, 1991). Also, Wong *et al.* (2000) found that the WHC for the *U. lactuca* seaweed at 37° C lower (9.71 g/g DW) with the result found at 37°C, but also comparable to that of some agricultural by-products (dietary fiber concentrates) (6.30–13.2 g/g DW) reported previously (Grigelmo-Miguel and Martin-Belloso, 1999). Furthermore, the Water Holding Capacity of the *C. racemosa* seaweed samples were also comparable to the Water Holding Capacity of some commercial dietary fiber rich supplements (Goñi and Martin-Carrón, 1998).

Seaweeds were rich in dietary fiber (>50% dry weight), particularly in the soluble form (Darcy-Vrillon, 1993; Mabeau *et al.*, 1993); Fleury *et al.* (1991) reported that the physico-chemical properties of seaweed powder could be assumed to reflect those of the present fibre. Moreover, since seaweed proteins are closely related to the cell wall polysaccharides (Fleurence, 1999), they may also play a role in the physicochemical properties, such as water holding.

Water exists in fiber in three forms: it is bound to the hydrophilic polysaccharides; it is held within the fiber matrix or it is trapped within the cell wall lumen. WHC, determined by the centrifugation method used in this study, represented all three types of water associated with the fiber (Fleury *et al.*, 1991). Apart from the different water holding ability in fiber, the differences in WHC and SWC among the seaweed samples might be attributed to the different protein conformations and the variations in the number and nature of the water binding sites on the protein molecules. In addition to chemical compositions, some physical properties, such as structure, particle size, porosity, pH, temperature, ionic strength, types of ions in solutions and density were important to the understanding of the different behaviors' of samples during hydration (Fleury *et al.*, 1991).

The results obtained in this study thus suggest the identified phytochemical compounds may be the bioactive constituents and the alga *C. racemosa* are proving to be an increasingly valuable reservoir of bioactive compounds of substantial medicinal merit.

CONCLUSION

The present study conclude that methanol solvent extract of *C. racemosa* shows a number of metabolites presence, of steroids, phenolic groups, saponins, tannin, flavonoids, terpenoids carbohydrates, reducing sugars, anthraquinones, Emodins coumarins, and xanthoproteins suggest that *C. racemosa* seaweed can be used as antimicrobial (anti-viral, anti-fungal and anti-bacterial), anti-parasitic, anti-inflammatory, antioxidant, antiallergenic, antithrombic, anticarcinogenic and anti-ulcer agents in the near future .

CONFLICT OF INTERESTS

The authors do not have any conflict of interests.

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REFERENCE

- Aguinaldo AM, El-Espeso, Guovara BQ, Nanoto M, 2005. Phytochemistry. In: Guevara BQ. (ed.) A guide book to plant screening phytochemical and biological. Manila: University of Santo Tomas.
- Ali, S.S., Kasoju, N., Luthra, A., Singh, A., Sharanabasava, H., Sahuand, A., Bora, U. 2008. Indian medicinal herbs as source of antioxidants. *Food Res. Int.*, 41: 1-15.
- Aliyu A.B., A.M. Musa, J.A. Oshanimi, H.A. Ibrahim and A.O. Oyewale, 2008. Phytochemical analyses and mineral elements composition of some medicinal plants of northern Nigeria . *Nig. Journ. Pharm. Sci.*, Vol. 7 (1), 119– 125.
- Anatharaman. P.G. Thirumaran., Balasubramanian T., 2007. Seaweed Farming: Alternative Livelihood. In Kannaiyan.S. and Venkataraman.(Eds). Biodiversity Conservation in Gulf of Manner Biosphere Reserve. National Biodiversity Authority, Chennai. 484pp.
- Aneiros A, Garateix A. 2004. Bioactive peptides from marine sources: pharmacological properties and isolation procedures. *J Chromatogr B*, 803:41–53.
- Anonymous, 1966. *Pharmacopoeia of India*. Ministry of Health, Government of India, Publication, New Delhi.
- Anonymous, 1985. *Pharmacopoeia of India*. Ministry of Health, Government of India Publication, New Delhi.
- Anonymous, 1996. *Pharmacopoeia of India*. Ministry of Health, Government of India Publication, New Delhi.
- Arts IC, Hollman PC. Polyphenols are disease risk in epidemiological studies, 2005. *Amer J Clin Nut*, 81: 317-325.
- Arun Kumar V and M Paridhavi. 2011. Evaluation of physiochemical parameters on the fruit of *Zanthoxylum limonella* Alston (FAMILY- RUTACEAE) *Pharmacie Globale* (IJCP), 11 (04).
- Athukorala, Y., Kim, K.N., Jeon, Y.J., 2006. Antiproliferative and antioxidant properties of an enzymatic hydrolysate from brown alga, *Ecklonia cava*. *Food Chem. Toxicol.* 44, 1065–1074.
- Athukorala, Y., Lee, K.W., Shahidi, F., Heu, M.S., Kim, H.T., Lee, J.S., Jeon, Y.J., 2003. Antioxidant efficacy of extracts of an edible red alga (*Grateloupia wlicina*) in linoleic acid and Wash Oil. *J. Food Lipid* 10, 313–327.
- Ayoola, GA, HAB Coker, SA Adesegun, AA Adepoju-Bello, K Obaweya, EC Ezennia, TO Atangbayila.2008. Phytochemical Screening and Antioxidant Activities of Some Selected Medicinal Plants Used for Malaria Therapy in Southwestern Nigeria. *Tropical Journal of Pharmaceutical Research*, 7 (3), 1019-1024.
- Barrow C, Shahidi F. 2008, Marine Nutraceuticals and Functional Foods. USA: CRC Press.
- Benjamin Jeya Rathna Kumar P and Christopher Patrick Kiladi S,2009. Preliminary Phytochemical and Pharmacognostic Studies of *Holoptelea integrifolia* Roxb. *Ethnobotanical Leaflets*, 13, 1222-31.

- Blunt JW, Copp BR, Hu WP, Munro MHG, Northcote PT, Prinsep MR, 2007. Marine natural products. *Nat Prod Rep*, 24: 31-86.
- Brindha P.,Sasikala B, and Purushothaman K.K.1991,BMEBR, 3(1):84-96
- Brown, J.E., Rice-Evans, C.A. 1998. Luteolin rich artichoke extract protects low density lipoprotein from oxidation in vitro. *Free Radical Res*, 29: 247-255.
- Chase, C.R. and Pratt, R.I. 1949. Fluorescence of powdered vegetable drugs with particular reference to development of, a system of identification. *J. Amer. Pharm. Ass.* 38: 324-331.
- Chitravadivu C., Manian S. and K. Kalaichelvi, 2009. Qualitative Analysis of Selected Medicinal Plants, Tamilnadu, India. *Middle-East Journal of Scientific Research*, 4 (3): 144-146.
- Cushnie TPT, Lamb AJ. 2005. Antimicrobial activity of flavonoids. *Int J Antimicrob Agents*,26 (5): 343-356.
- De Sousa RR, Queiroz KC, Souza AC, Gurgueira SA, Augusto AC, Miranda MA, 2007. Phosphoprotein levels, MAPK activities and NFkappaB expression are affected by fisetin. *J Enzyme Inhib Med Chem*, 22 (4): 439-444.
- Del-Rio, A., Obdululio, B.G., Casfallo, J., Main, F.G., Ortuno, A. 1997. Uses and properties of citrus flavonoids. *J. Agric. Food Chem.*, 45: 4505-4515.
- De-Lucca A, Cleveland T, Rajasekara K, Boue S, Brown R. 2005.Fungal properties of CAY-1, a plant saponin, for emerging fungal pathogens. 45th inter science conference in antimicrobial agents and chemotherapy abstract.; p.180.
- Devmurari V. P. and Jivani N. P. 2010. Phytochemical screening and antibacterial activity of ethanolic extract of *Artemisia Nilagirica*. *Annals of Biological Research*, 1 (1), 10-14.
- Duan, X.J., Zhang, W.W., Li, X.M., Wang, B.G., 2006. Evaluation of antioxidant property of extract and fractions obtained from a red alga, *Polysiphonia urceolata*. *Food Chem.* 95, 37-43.
- Duygu Dilek, Abel U. Udoh, Tulay (Baykal) Ozer, Aydin Akbulut, Ilkay (Acikgoz) Erkaya, Kazim Yildiz and Deniz Guler, 2012. Fourier transform infrared (FTIR) spectroscopy for identification of *Chlorella vulgaris* Beijerinck 1890 and *Scenedesmus obliquus* (Turpin) Kützing 1833. *African Journal of Biotechnology* , 11(16), 3817-3824.
- Edeoga, H.O, D. E. Okwu and B.O Mbaebie. 2005. Phytochemical constituents of some Nigerian medicinal Plants. *African Journal of Biotechnology* Vol. 4 (7), 685-688.
- Elena, M., Francisco, Y., and Erickson, K.L., 2001. Mailiohydrin, a Cytotoxic Chamigrene Dibromohydrin from a Phillippine *Laurencia* Species, *J. Nat. Prod.*, Vol. 64, No. 6, pp. 790-791.
- Eluvakkal T, Sivakuamr SR, Arunkumar K, 2010. Fucoidan in some Indian brown seaweeds found along the coast of Gulf of Mannar. *Int J Botany*, 6(2): 176-181.
- Faulkner DJ, 2002. Marine Natural Products. *Nat Prod Rep*, 19:1-48.
- Faulkner, D. J. 2001. Marine Natural Products. *Nat. Prod. Rep.*, 18: 1-49.
- Fleurence, J., 1999. Seaweed proteins: biochemical nutritional aspects and potential uses. *Trends in Food Science and Technology.*, 10(1): 25-28.
- Fleury, N., and Lahaye, M., 1991. Chemical and physico-chemical characterization of fibers from *Laminaria digitata* (Kombu Breton): a physiological approach. *Journal of Science and Food Agriculture.*, 55: 389-400
- Fuesetani, N. 2000. Drugs from the sea. Basel: *Karger*, p. 1-5.
- George F, Zohar Kerem, Harinder PSM, Klaus Becker. 2002. The biological action of saponins in animal systems: a review. *Brit J Nutr*; 88(6): 587-605.
- Giancaspro Gabriel I, 2007. Dietary Supplements (DSB): Botanicals, USP27-NF22 ,1993, British Pharmacopeia , Forum : Volume No. 26(6) 1594.
- Hamad H. Hasan, Ibrahim H. Habib1, Mariam. H. Gonaïd and Mojahidul Islam. 2011. Comparative phytochemical and antimicrobial investigation of some plants growing in Al Jabal Al-Akhdar. *J. Nat. Prod. Plant Resour*, 1 (1):15-23.
- Han, X., Shen, T., Lou, H. 2007. Dietry polyphenols and their biological significance. *Int. J. Mol. Sci.*, 950-988.
- Harborne JB.1998. Phytochemical methods. London: Chapman and Hall.
- Heo, S.J., Park, E.J., Lee, K.W., Jeon, Y.J., 2005. Antioxidant activities of enzymatic extracts from brown seaweeds. *Bioresour. Technol.* 96, 1613-1623.

- Holdt, S. L. and Kraan, S., 2011. Bioactive compounds in seaweed: Functional food applications and legislation. *J. Appl. Phycol.*, 23: 543–598.
- Iyengar, M.O.P. 1933. On the formation of gametes in a *Caulerpa*. *J. Indian Bot. Soc.*, XII : 325.
- Janakiraman N, Sahaya Sathish S And Johnson M, 2011. UV-Vis and FTIR Spectroscopic Studies on *Peristrophe bicalyculata* (Retz.) Nees. *Asian J Pharm Clin Res*, 4, (4) 125-129.
- Jayasree, N.B, Aneesh.T.P, Visakh Prabhakar and R.Anandan . 2012. GC-MS, HPLC and AAS analysis of Fatty Acids, Amino Acids And Minerals in Red Algae *Amphiroa anceps*. *Int J Pharm Pharm Sci, Vol 4, Suppl 1*, 187-190.
- Jeeva Solomon, Johnson Marimuthu Antonisamy, Cosman Domettila, Babu Anantham, Mony Mahesh 2012. Preliminary phytochemical studies on some selected seaweeds from Gulf of Mannar, India. *Asian Pacific Journal of Tropical Biomedicine*, S30-S33.
- Jeeva Solomon, Johnson Marimuthu Antonisamy. 2012. Anti-bacterial and phytochemical studies on methanolic extracts of *Begonia floccifera* Bedd. Flower . *Asian Pacific Journal of Tropical Biomedicine*, S151-S154.
- Johnson Marimuthu Antonisamy, Petchiammal Essakimuthu, Janakiraman Narayanan, Babu Anantham, Renisheya Joy Jeba Malar Tharmaraj, Sivaraman Arumugam, 2012. Phytochemical characterization of brown seaweed *Sargassum wightii*. *Asian Pacific Journal of Tropical Disease*, S109-S113.
- Johnson, M, Maridass, M and Irudayaraj V, 2008. Preliminary Phytochemical and Anti-Bacterial Studies on *Passiflora edulis*. *Ethnobotanical Leaflets*, 12, 425-432.
- Just, M.J., Recio, M.C., Giner, R.M., Cueller, M.U., Manez, S., Billia, A.R., Rios, J.L. 1998. Antiinflammatory activity of unusual lupine saponins from *Bupleurum fruticosens*, 64: 404-407.
- Kapoor LD, Singh A, Kapoor SL and Shrivastava SN, 1969. Survey of Indian medicinal plants for saponins, alkaloids and flavonoids. *Lloydia*, 32: 297-302.
- Khandalwal KR, 2008. Practical Pharmacognosy- Techniques and Experiments. Nirali Prakashan. 19th ed, 149-156.
- Kim SJ, Woo S, Yun H, Yum S, Choi E, Do JR, 2005. Total phenolic contents and biological activities of Korean seaweed extracts. *Food Sci Biotechnol*, 14:798–802.
- Kim SK, Wijesekara I, 2010. Development and biological activities of marine-derived bioactive peptides: a review. *J Functional Foods*, 2:1–9.
- Kim, J.H., Hudson, J.B., Huang, A.M., Bannistes, K., Jin, H., Choi, T.J., Towers, G.H.N., Hong, Y.K., and DeWreede, R.E., 1997. Biological Activities of Seaweed Extracts from British Columbia, Canada, and Korea. I. Antiviral Activity," *Can. J. Bot. Rev. Can. Bot.*, Vol. 75, No. 10, pp. 1656-1660.
- Kokate C K, 1997. Practical Pharmacognosy. 4th ed. Vallabh Prakashan.; 53, 123-124,127.
- Kokate CK, Purohit AP, Gokhale SB, 2005. Pharmacognosy, 39th Edition, Nirali Prakashan, Pune, 607-611.
- Kokoshi, C.J., Kokoshi, R.J. and Slama, F.J. 1958. Fluorescence of powdered vegetable Drug under U.V.Radiation. *Sci. Ed. J. Am. Pharm. Associ.* 48 (10): 715-717.
- Kolodziej H, Kiderlen AF. 2005. Antileishmanial activity and immune modulatory effects of tannins and related compounds on *Leishmania* parasitised RAW 264.7 cells. *Phytochem*, 66 (17): 2056-2071.
- Krings, U., Berger, R.G. 2001. Antioxidant activity of roasted foods. *Food Chem.*, 72: 223-229.
- Kuda, T., Kunii, T., Goto, H., Suzuki, T., Yano, T., 2007. Varieties of antioxidant and antibacterial properties of *Ecklonia stolonifera* and *Ecklonia kurome* products harvested and processed in the Noto peninsula, Japan. *Food Chem.* 103, 900–905.
- Li Yong-Xin, Isuru Wijesekara, Yong Li, Se-Kwon Kim, 2011. Phlorotannins as bioactive agents from brown algae. *Process Biochemistry*, 46, 2219–2224.
- Mabeau, S., Fleurence, J., 1993. Seaweed in food products: biochemical and nutritional aspects. *Trends in Food Science and Technology*, 4: 103–107.
- Majaw S. and J. Moirangthem. 2009. Qualitative and Quantitative Analysis of *Clerodendron colebrookianum* Walp. Leaves and *Zingiber cassumunar* Roxb. Rhizomes. *Ethnobotanical Leaflets* 13:578-89.

- Mandal P, Sinha Babu SP, Mandal NC, 2005. Antimicrobial activity of Saponins from *Acacia auriculiformis*. *Fitoterapia*, 76(5): 462-565.
- Manilal, A., S. Sujith, B. Sabarathnam, G.S.Kiran, J. Selvin, C. Shakir and A. P.Lipton., 2010. Bioactivity of the red alga *Asparagopsis taxiformis* collected from the south-western coast of India. *Brazilian J. Oceanography*, 58(2): 93-100.
- Manilal, A., S. Sujith, G. S. Kiran, J. Selvin, C.Shakir, R. Gandhimathi and A. P. Lipton .,2009a. Antimicrobial potential and seasonality of red algae collected from the Southwest coast of India tested against shrimp, human and phytopathogens. *Ann. Microbiol.*, 59(2): 207-219.
- Manilal, A., Sujith, S., Kiran, G. S., Selvin, J., Shakir, and C., Gandhimathi, R., 2009b.Bio-potentials of seaweeds collected from southwest coast of India. *Journal of Marine Science and Technology*, 17, 67-73.
- Manjunatha BK. Antibacterial activity of *Pterocarpus santalinus*, 2006. *Ind J Pharm Sci*, 68(1): 115-116.
- Maridass M. 2010. Survey of Phytochemical Diversity of Secondary Metabolism in Selected Wild Medicinal Plants. *Ethnobotanical Leaflets*, 14, 616-25.
- Maridass M., M.I. Zahir Hussain and G. Raju 2008b. Phytochemical Survey of Orchids in the Tirunelveli Hills of South India. *Ethnobotanical Leaflets*, 12, 705-12.
- Maridass M., S. Ganthikumar and G. Raju, 2008a. Preliminary Phytochemical Analysis of *Diospyros Species*. *Ethnobotanical Leaflets*, 12: 868-72.
- Marjorie, C. 1996. Plant products as antimicrobial agents. *Clinical Microbiol. Rev.*, 12: 564-582.
- Meenakshi S, Manicka GD, Tamilmozhi S, Arumugam M, Balasubramanian T. 2009. Total flavanoid and *in vitro* antioxidant activity of two seaweeds of Rameshwaram Coast, *Global J. Pharmacol*, 3: 59-62.
- Meenakshi Selvaraju Shanmugam Umayaparvathi, Muthuvel Arumugam, Thangavel Balasubramanian, 2012. *In vitro* antioxidant properties and FTIR analysis of two seaweeds of Gulf of Mannar. *Asian Pacific Journal of Tropical Biomedicine*, S66-S70.
- Mendis Eresha and Se-Kwon Kim, 2011. Present and Future Prospects of Seaweeds in Developing Functional Foods. *Advances in Food and Nutrition Research*, Volume 64 1-15.
- Mensah J.K., R. Okoli , A. A. Turay, and E.A. Ogie-Odia, 2009. Phytochemical Analysis of Medicinal Plants Used for the Management of Hypertension by Esan people of Edo State, Nigeria. *Ethnobotanical Leaflets*, 13, 1273-87.
- Mohanta TK, Patra JK, Rath SK, Pal DK, Thatoi HN.2007. Evaluation of antimicrobial activity and phytochemical screening of oils and nuts of *Semicarpus anacardium* L.f. *Sci Res Essay*; 2(11): 486-490.
- Naga Deepthi, Bolla, Praveen kumar and Bhogavalli, 2010. Preliminary phytochemical screening and antibacterial studies of the flowers of *Antigonon leptopus* . *Annals of Biological Research*, 1 (4) : 229-233.
- Nazish I, Kaskoos R A, Mir S R, Amin S and Ali M; 2009. Preliminary pharmacognostical standardization of *Ruta graveolens* L. Aerial Parts. *Research Journal of Medicinal Plant*; 3(2):41-44.
- Nwosu F, Morris J, Lund VA, Stewart D, Ross HA, McDougall GJ, 2011. Anti-proliferative and potential anti-diabetic effects of phenolic-rich extracts from edible marine algae. *Food Chem*, 126:1006-12.
- Okai, Y., Higashi, O.K., Ishizaka, S., and Yamashita, U., 1997. Enhancing Effect of Polysaccharide from an Edible Brown Alga, *Hijikia fusiforme* (Hijiki), on Release of Tumour Necrosis Factor Alpha from Macrophages of Endotoxin Nonresponder C3H/HeJ Mice," *Nutr.Cancer*, Vol. 27, No. 1, pp. 74-79 .
- Okhale, Samuel Ehiabhi, Odiniya, Emmanuel Odiniya and Kunle, Oluyemisi Florence 2010. Preliminary Phytochemical and Pharmacognostical Investigation of Pediatrics Antimalarial *Laggera pterodonta* (DC) Sch. Bip.: Asteraceae of Nigerian Origin. *Ethnobotanical Leaflets*, 14: 457-66.
- Okwu, D.E. 2001. Evaluation of chemical composition of medicinal plants belonging to Euphorbiaceae. *Pak Vet. J*,14: 160-162.

- Okwu, D.E. 2004. Phytochemicals and vitamin content of indigenous species of southeastern Nigeria. *J.Sustain. Agric. Environ.*, 6(1): 30-37.
- Paris, R. and H. Moyses, 1969. *Precis de matiere medicinale*. Paris: Masson.
- Plaza M, Cifuentes A, Ibanez E, 2008. In the search of new functional food ingredients from algae. *Trends Food Sci Technol*, 19:31-9.
- Pratt RT, Chase ER, 1949. Fluorescence powder vegetable drugs in particular to development system of identification. *J. Am. Pharm. Assoc*, 38: 324-331.
- Premila, J.C., Raviraja, N.S., and Sridhar, K.R., 1996. Antimicrobial Activity of Some Marine Algae of Southwest Coast of India, *Indian J. Mar. Sci.*, Vol. 26, No. 2, pp. 201-205.
- Quality control methods for medicinal plant materials. WHO, Geneva. A. I. T. B. S. Publishers & Distributors (Regd.) Delhi. 51. 2002; 10, 30, 45, 46.
- Rafia Rasool, Bashir A Ganai, Seema Akbar, Azra N Kamili and Akbar Masood. 2010. Phytochemical screening of *Prunella vulgaris* L. an important medicinal plant of Kashmir. *Pak. J. Pharm. Sci.*, Vol.23, No.4,399-402.
- Rajasulochana P, Dhamotharan R, Krishnamoorthy P. 2009. Primary Phytochemical Analysis of *Kappaphycus* Sp. *Journal of American Science*, 5(2): 91-96.
- Raquel, F.E. 2007. Bacterial lipid composition and antimicrobial efficacy of cationic steroid compounds. *Biochemica et Biophysica Acta*, 2500-2509.
- Rievere C, Van Nguyen JH, Pieters L, Dejaegher B, Heyden YV, Minh CV 2009. Polyphenols isolated from antiradical extracts of *Mallotus metcalfeanus*. *Phytochemistry*, 70: 86-94.
- Rizk, A.M., 1982. *Fitoterapia*, 52: 35-4234.
- Sakthivel, 2007. Seaweed Cultivation: A profitable venture for the Economic Rehabilitation of coastal Poor. In Kannaiyan .S. and Venkataraman.(eds). Biodiversity Conservation in Gulf of Manner Biosphere Reserve. National Biodiversity Authority, Chennai. 484pp.
- Salah, N., Miller, N.J., Pagange, G., Tijburg, L., Bolwell, G.P, Rice, E., Evans, C. 1995. Polyphenolic flavonoids as scavenger of aqueous phase radicals as chai breaking antioxidant. *Arc. Biochem. Broph.*, 2: 339-346.
- Savithamma, M. Linga Rao and D. Suhrulatha. 2011. Screening of Medicinal Plants for Secondary Metabolites. *Middle-East Journal of Scientific Research*. 8 (3): 579-584,.
- Scalbert A, Manach C, Morand C, Remesy C, Jimenez L. 2005. Dietary polyphenols and the prevention of diseases. *Cri Rev Food Sci Nutr*, 45: 287-306.
- Selvin Joseph and Aaron Premnath Lipton 2004. Biopotentials Of *Ulva Fasciata* And *Hypnea Musciformis* Collected From The Peninsular Coast Of India. *Journal of Marine Science and Technology*, Vol. 12, No. 1, pp. 1-6.
- Selvin, J. and Lipton, A.P., 2004. Biopotentials of Secondary Metabolites Isolated from Marine Sponges. *Hydrobiologia*, Vol. 513, 231-238 .
- Singh, R., Singh, S.K., Arora, S. 2007. Evaluation of antioxidant potential of ethyl acetate extract/fractions of *Acacia auriculiformis* A. Cunn. *Fod Chem. Toxicol.*, 45: 1216-1223.
- Smit AJ, 2004. Medicinal and pharmaceutical uses of seaweed natural products: a review. *J Appl Phycol*, 16:245-62.
- Smolenski SJ, Silinis H, Farnsworth NR ,1974. Alkaloids Screening. *V. Lloydia*, 37: 506-536.
- Sodipo, O.A., Akinyi, J.A., Ogunbamosu, J.U. 2000. Studies on certain characteristics of extracts of bark of *Pansinystalia macruceras* (K schemp) picrre Exbeille. *Global J. Pure Appl. Sci*, 6: 83-87.
- Sunita Dalal, Sudhir K Kataria, KV Sastry and SVS Rana, 2010. Phytochemical Screening of Methanolic Extract and Antibacterial Activity of Active Principles of Hepatoprotective Herb, *Eclipta alba*, *Ethnobotanical Leaflets*, 14, 248-58.
- Trease GE and Evans WC .1996. Pharmacognsy. 11th edn. Macmillian Publishers ,832pp .
- Ujowundu C. O., Okafor, O. E, Agha ,C., L. A. Nwaogu, K. O. Igwe and C. U. Igwe, 2010. Phytochemical and chemical composition of *Combretum zenkeri* leaves. *Journal of Medicinal Plants Research*, Vol. 4(10), 965-968.
- Usha Veerachari and A. K. Bopaiah, 2011. Preliminary phytochemical evaluation of the leaf extract of five *Cassia Species*. *J. Chem. Pharm. Res.*, 3(5):574-583.

- Venkata Ratnam, K and R.R. Venkata Raju, 2009. Preliminary Phytochemical and Antimicrobial Properties of *Pueraria tuberosa* (Willd.) DC: A Potential Medicinal Plant. *Ethnobotanical Leaflets*, 13, 1051-59.
- Venkatesh R. Shanthi S, Rajapandian K, Elamathi S. Thenmozhi S. and Radha N, 2011. Preliminary Study on Antixanthomonas Activity, Phytochemical analysis, and Characterization of antimicrobial compounds from *Kappaphycus alvarezii*. *Asian J Pharm Clin Res*, 4, (3), 46-51.
- Wang, T., Jonsdottir, R., Olafsdottir, G., 2009. Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds. *Food Chem.* 116, 240-248.
- Wijesekara I, Kim SK, 2010. Angiotensin-I-converting enzyme (ACE) inhibitors from marine resources. *Mar Drugs*, 8:1080-93.
- Wijesekara I, Pangestuti R, Kim SK, 2011. Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. *Carb Polymer*, 84:14-21.
- Wijesekara I, Yoon NY, Kim SK, 2010. Phlorotannins from *Ecklonia cava* (Phaeophyceae): biological activities and potential health benefits. *Biofactors*, 36:408-14.
- Xu R, Zhao W, Xu J, Shao B, Qin G. 1996. Studies on bioactive saponins from Chinese medicinal plants. *Adv Exp Med & Biol*, 404, 371-382.
- Yadav RNS and Munin Agarwala, 2011. Phytochemical analysis of some medicinal plants *Journal of Phytology*, 3(12): 10-14.
- Yamunadevi Mariswamy, Wesely Edward Gnanaraj, Johnson Marimuthu Antonisamy, 2012. FTIR Spectroscopic Studies on *Aerva lanata* (L.) Juss. Ex Schult. *Asian J Pharm Clin Res*, 5, (2), 82-86.
- Yan, X.J., Li, X.C., Zhou, C.X., Fan, X., 1996. Prevention of fish oil rancidity by phlorotannins from *Sargassum kjellmanianum*. *J. Appl. Phycol.* 8, 201-203.